



Bioconjugates

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Conditions

1. 5-7 lectures plus substantial homework
2. Participation at lectures > 70 %
3. Examination: 25-30 min presentation based on literature

Website:

<http://szerves.chem.elte.hu/oktatas/ea/Hudecz/index.htm#biokonjugatumok>

Email: ferenc.hudecz@ttk.elte.hu

Outline: Protein bioconjugates

1. Historical background

2. Functional groups of proteins/glycoproteins

N-nucleophiles: $-\text{NH}_2$, imidazole, indole, guanidino

S-nucleophiles: $-\text{SH}$, $\text{CH}_2-\text{S}-\text{CH}_3$

O-nucleophile: $-\text{OH}$

O/C-nucleophiles: $-\text{CHO}$, $-\text{COOH}$, $-\text{CONH}_2$

3. Creation of reactive groups

- Limited reactivity (e.g. $-\text{OH}$ vs. $-\text{CHO}$)

- Improved selectivity (e.g. $-\text{NH}_2$ vs. $-\text{SH}$)

- Space considerations

- Convenient chemistry (e.g. $-\text{COOH}$ vs. $-\text{NH}_2$)

Introduction

Transformation

4. Detection of reactive groups

sensitive

quantitative

quick

small sample

Destructive

Non-destructive

5. Conjugation

- Chemical synthesis

- Enzymatic synthesis (e.g. $-\text{NH}_2$ vs. $-\text{SH}$)

- Gene technology

6. Analysis of conjugates

Purification

Structure determination

Introduction

Design of bioconjugates

Why?

Synthetic antigens or drug targeting

What?

Peptide epitope, drug, reporter molecules

With What?

Protein, DNA, liposome

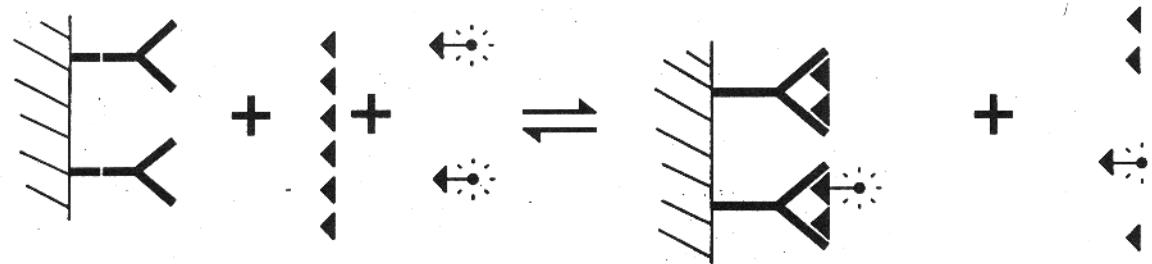
How?

Covalent bond

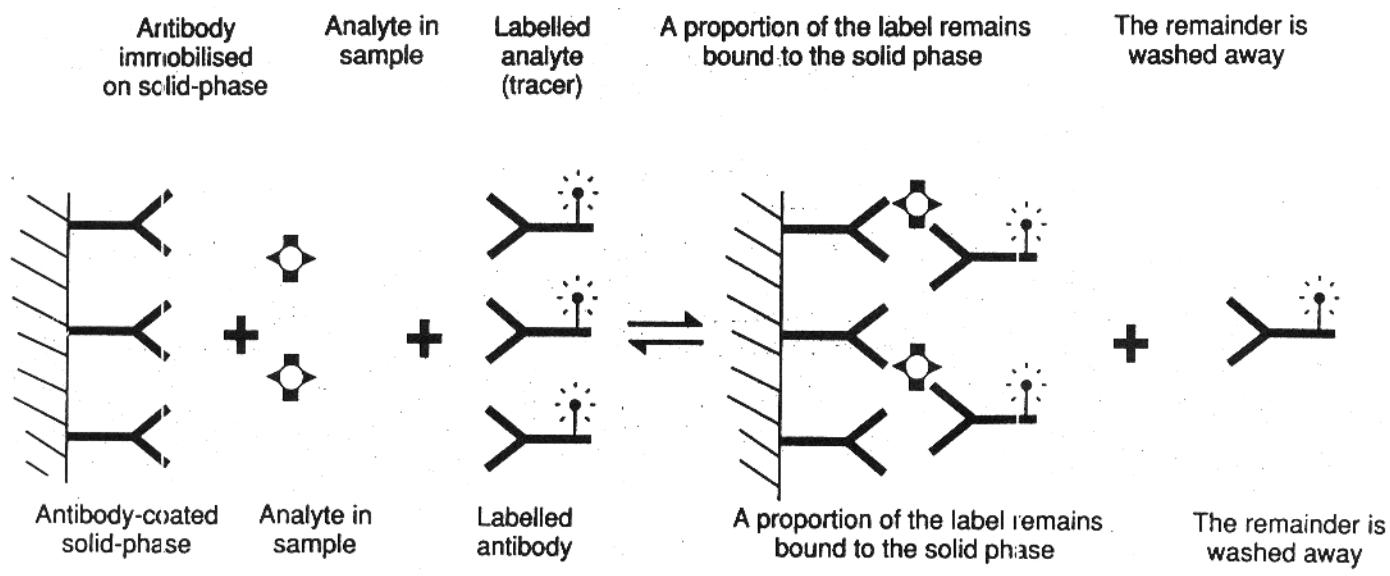
Introduction

Application of bioconjugates: immunoassays

a) competitive

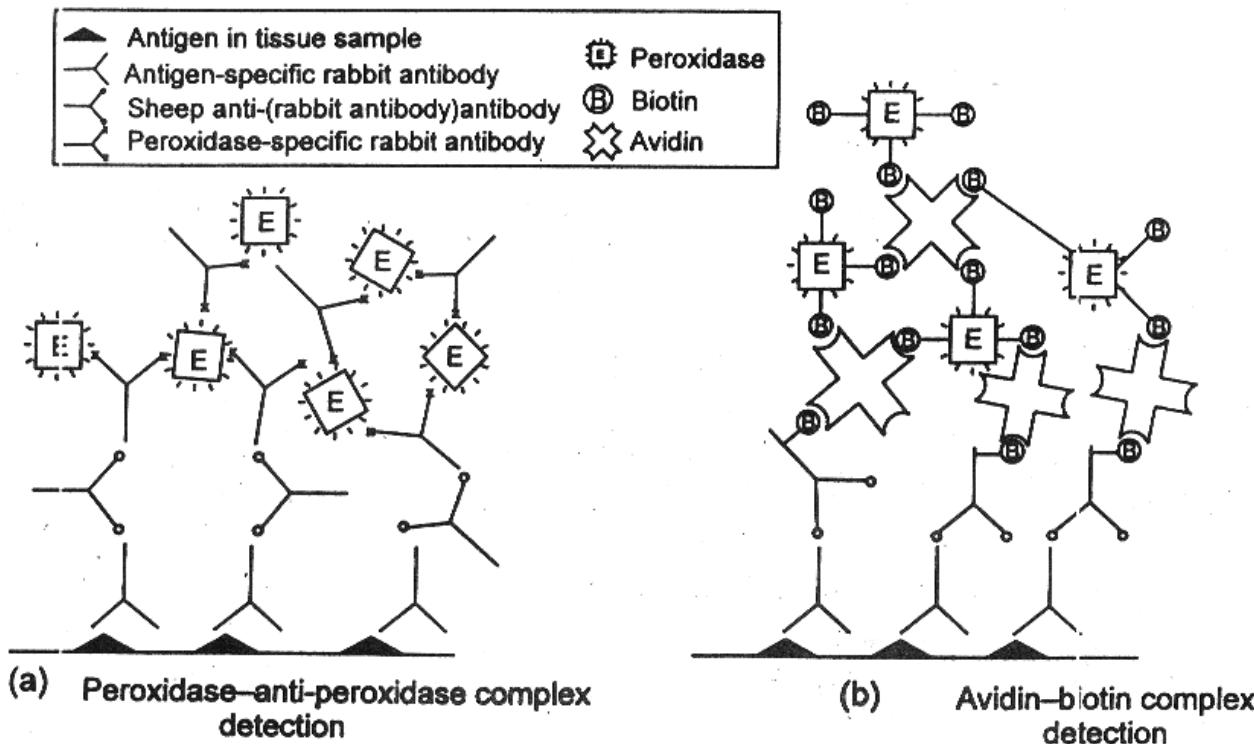


b) direct



Introduction

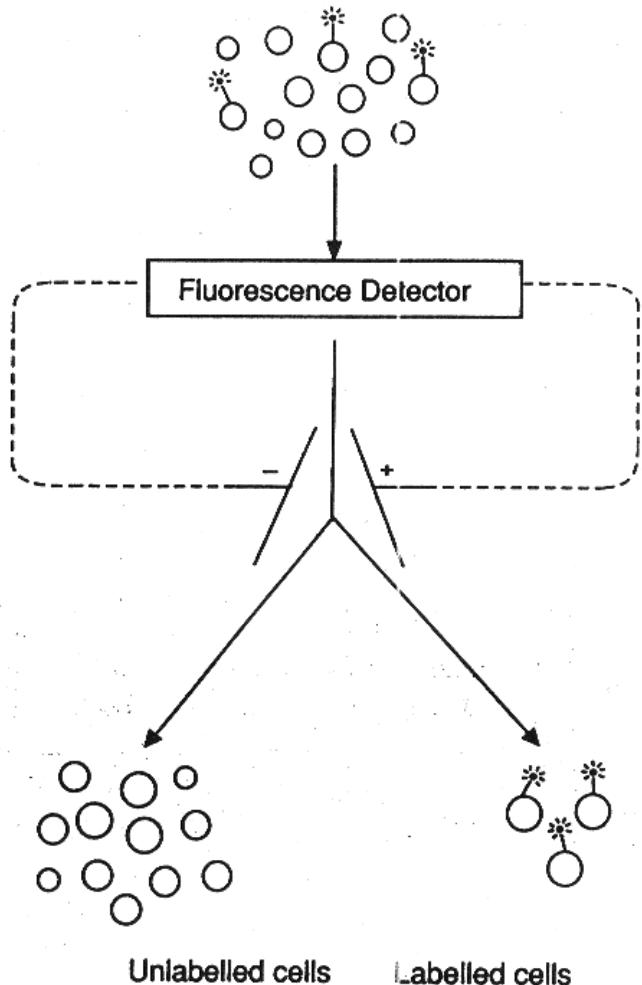
Application of bioconjugates: histo- and cytochemistry



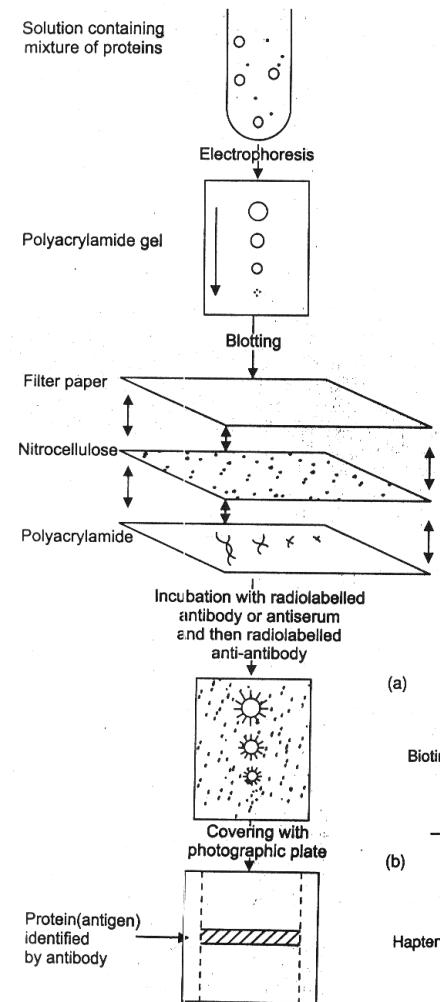
Introduction

Application of bioconjugates

FACS analysis

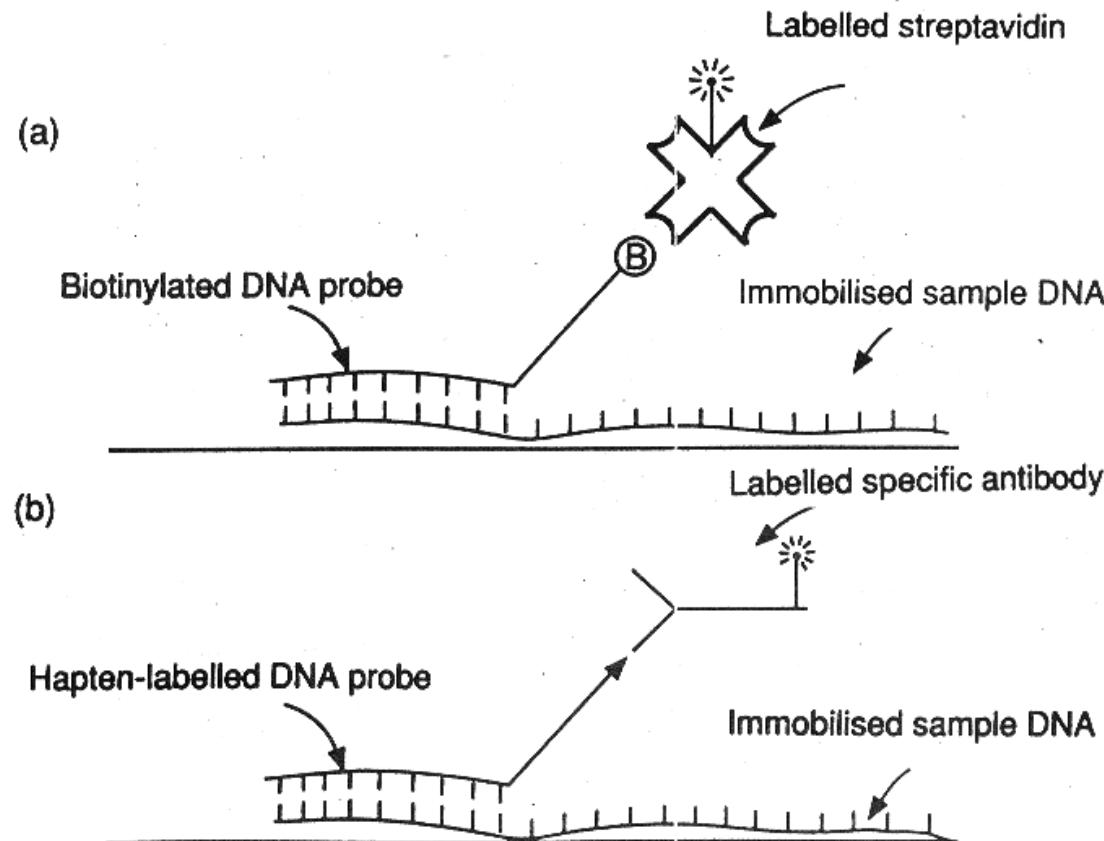


Western blotting

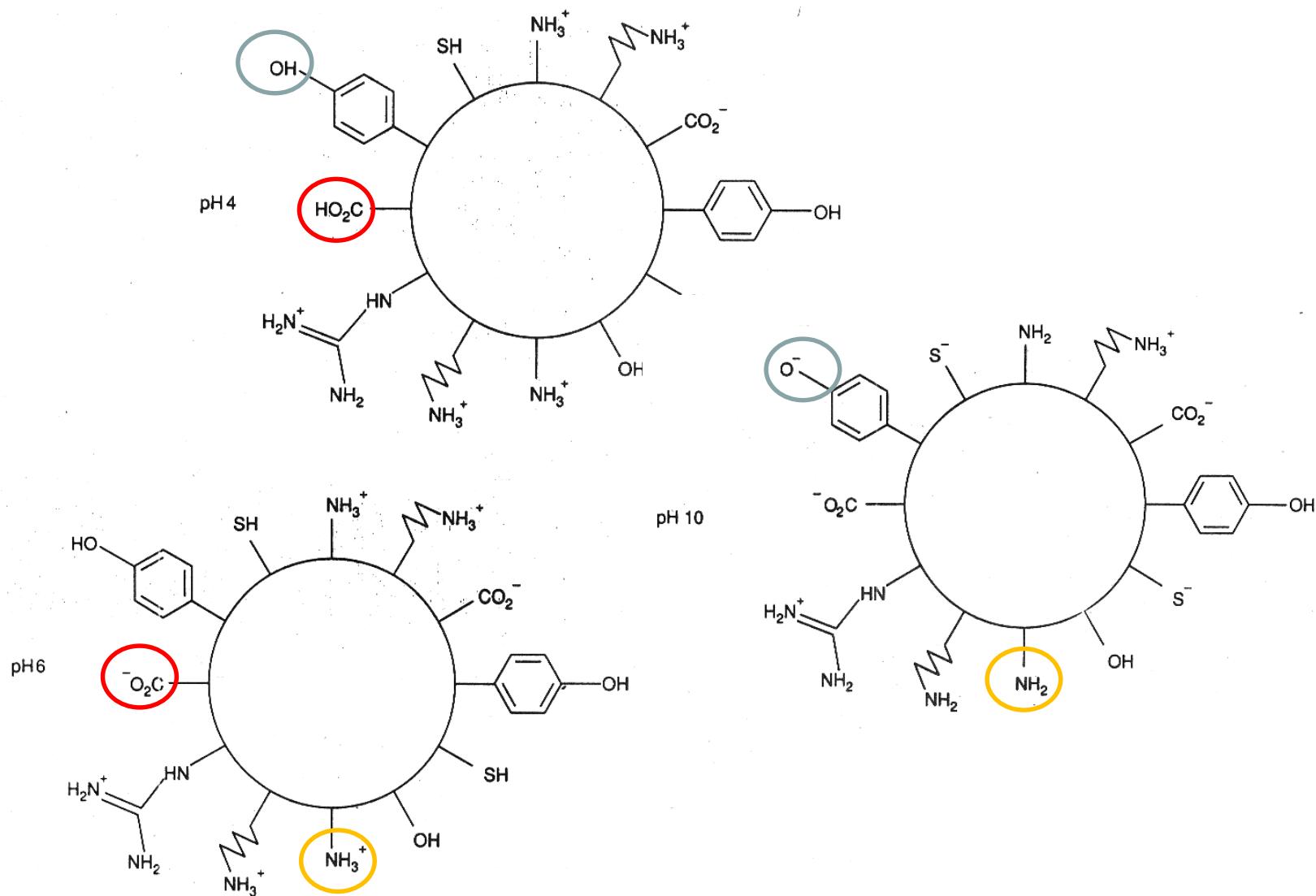


Introduction

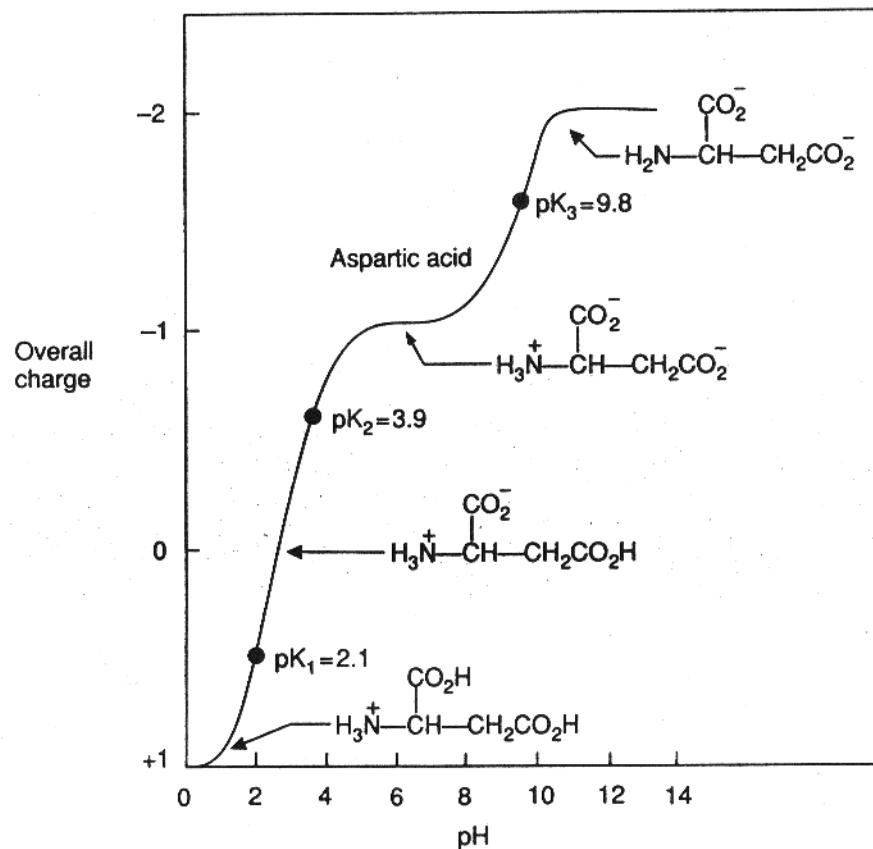
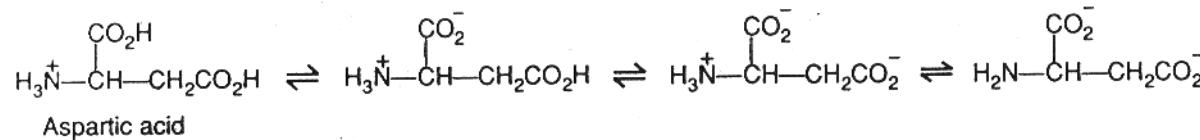
Application of bioconjugates: DNA detection



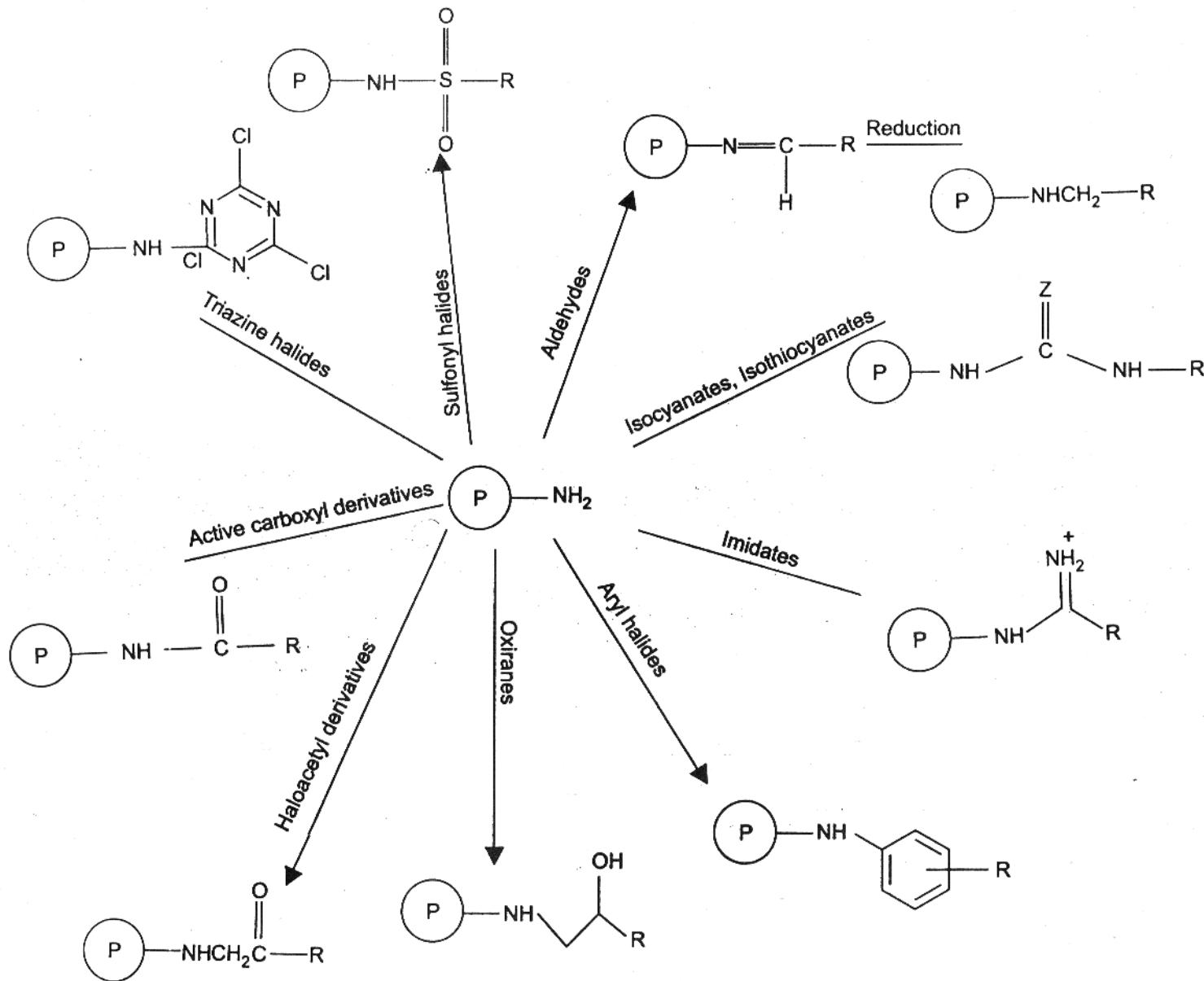
Ionisation status of proteins as function of pH



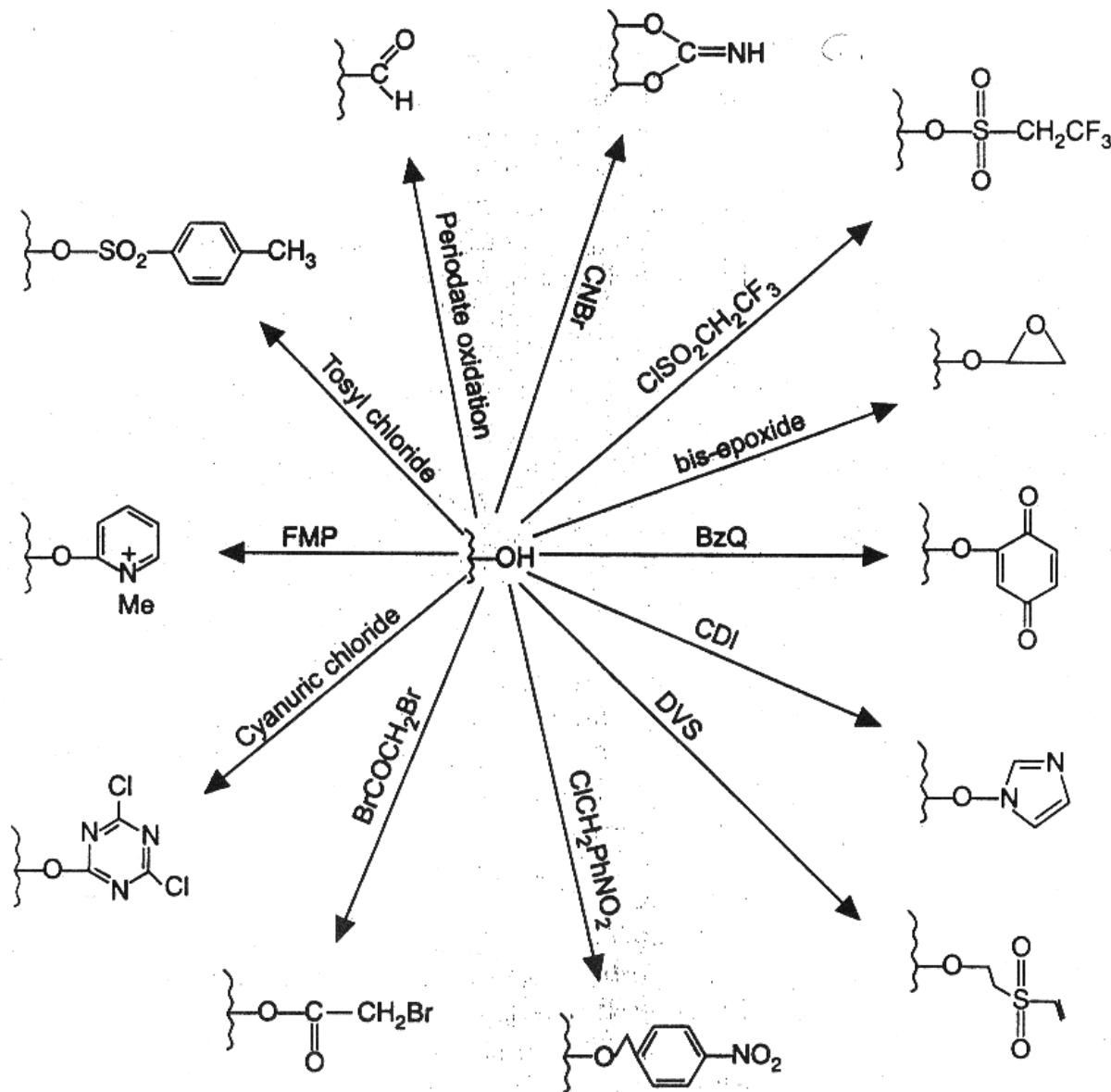
Ionisation of aspartic acid



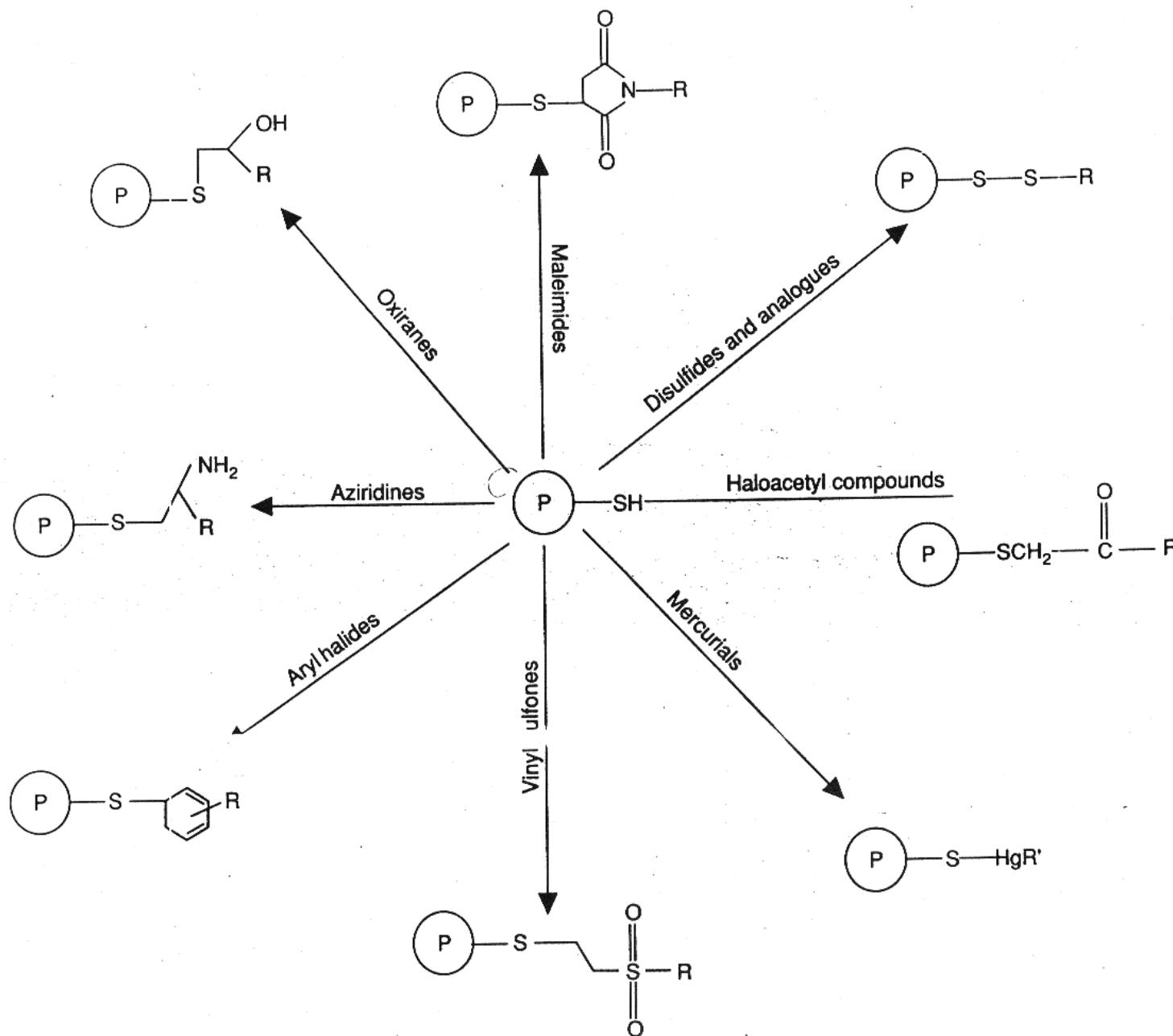
Reactivity and transformation of amino group



Reactivity and transformation of hydroxyl group



Reactivity and transformation of thiol group



Historical background

1) Establishment of protein structure - function relationship (1925 -

- Solubility

Sumner, J.B., Graham V.A.: The nature of **insoluble** creatine Proc Soc Exp Biol Med 22 504 (1925)

- Identification of amino acid side chain(s) necessary for the protein function

Olcott, H.S., Fraenkel-Conrat, H.:

Specific group reagents for proteins Chem Rev 41 151 (1947)

Herriot, R.M.:

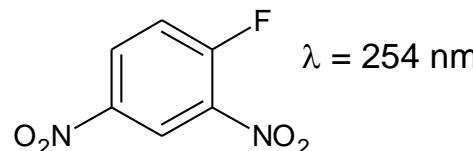
Reactions of native proteins with chemical reagents

Adv Prot Chem 3 161 (1947)

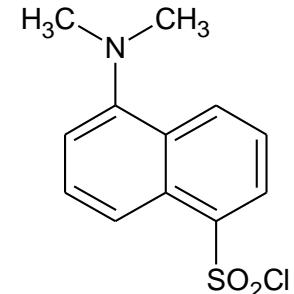
2) Structural studies

□ Proteins

■ Determination of N-terminal amino acid



$\lambda = 254 \text{ nm}$

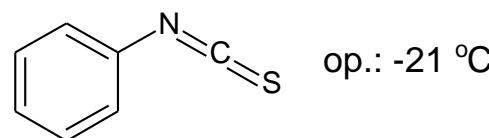


$\lambda_s = 360 \text{ nm}$

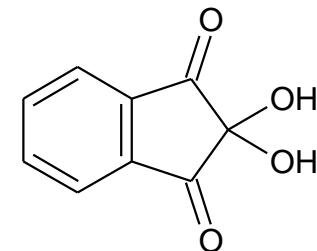
$\lambda_e = 480 \text{ nm}$

■ Sequencing (1956)

Edman, P., Begg, G.: A protein sequenator *Eur J Biochem* 1 80 (1967)



op.: -21 °C

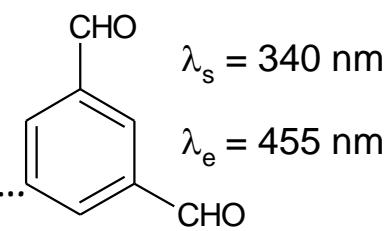


■ Determination of amino acid composition (1960)

Moore, S., Stein, W.H.:

Chromatographic determination of amino acids ...

Methods in Enzymol 6 819 (1963)

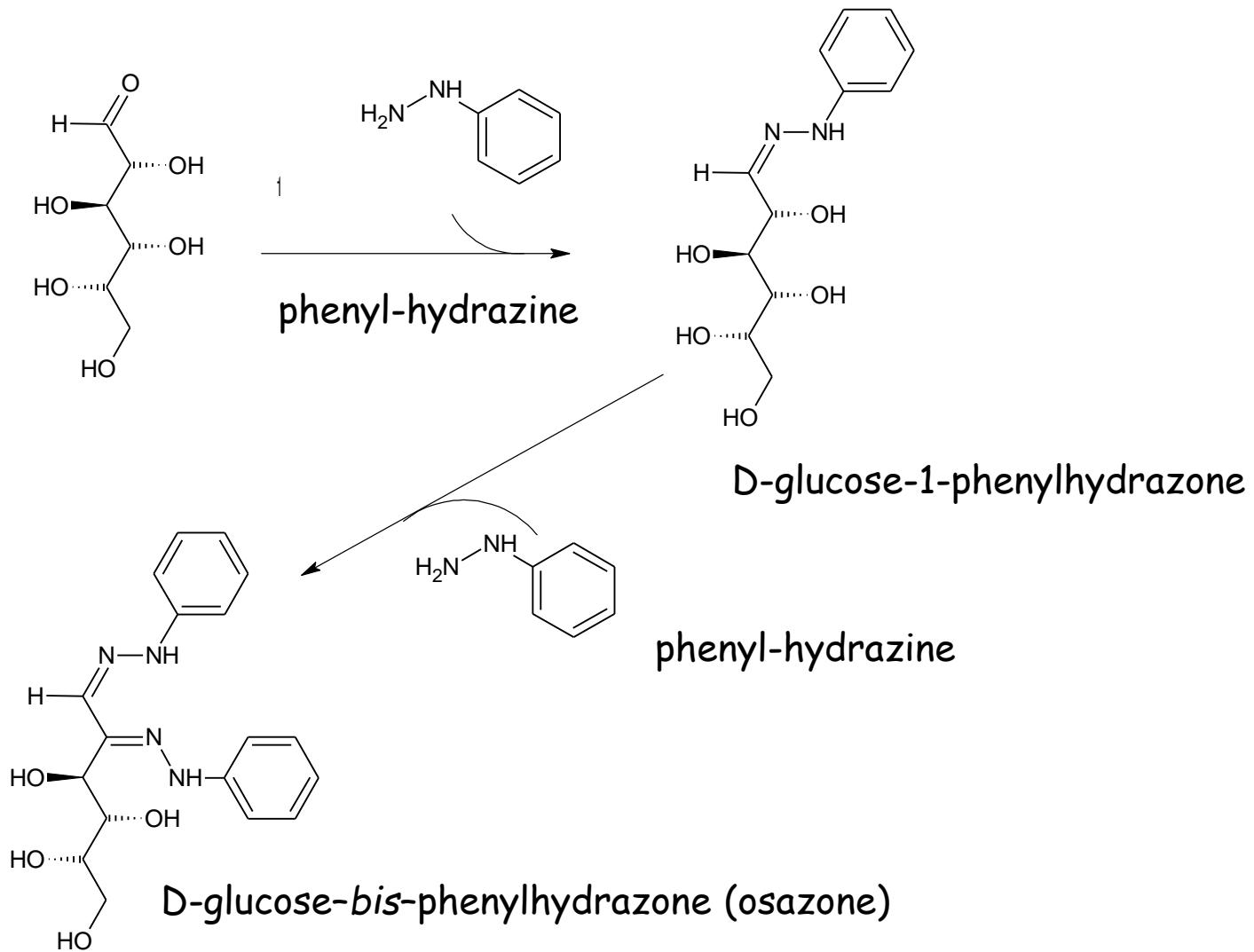


$\lambda_s = 340 \text{ nm}$

$\lambda_e = 455 \text{ nm}$

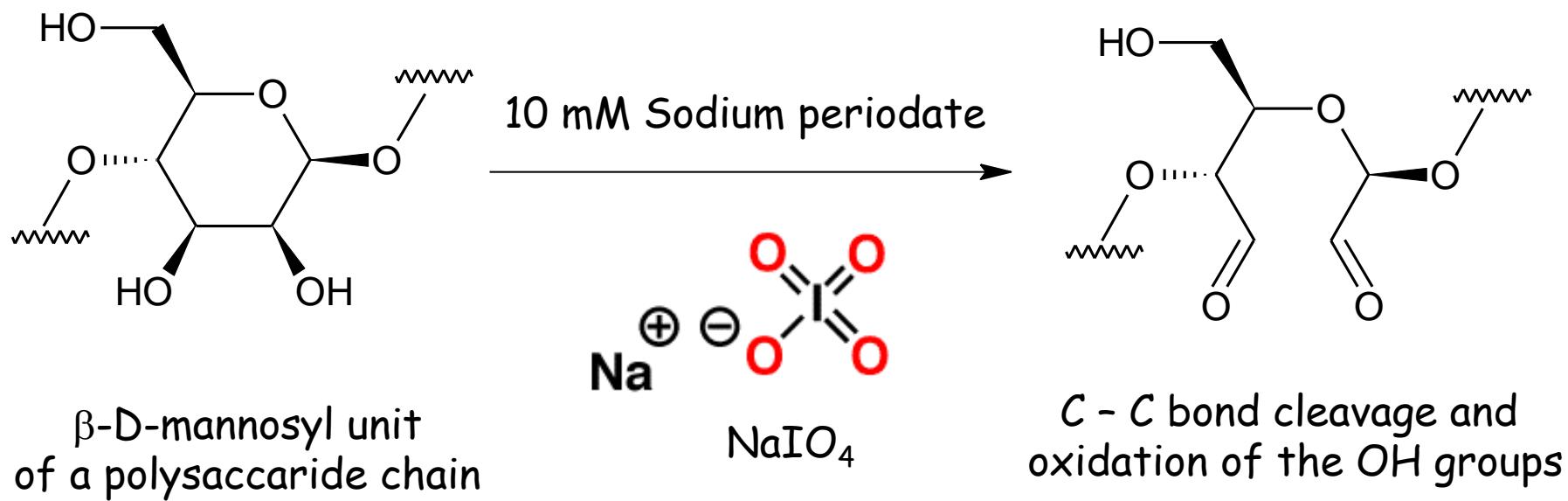
□ Carbohydrates

- E. Fischer (1884) Reducing monosaccharides



□ Carbohydrates

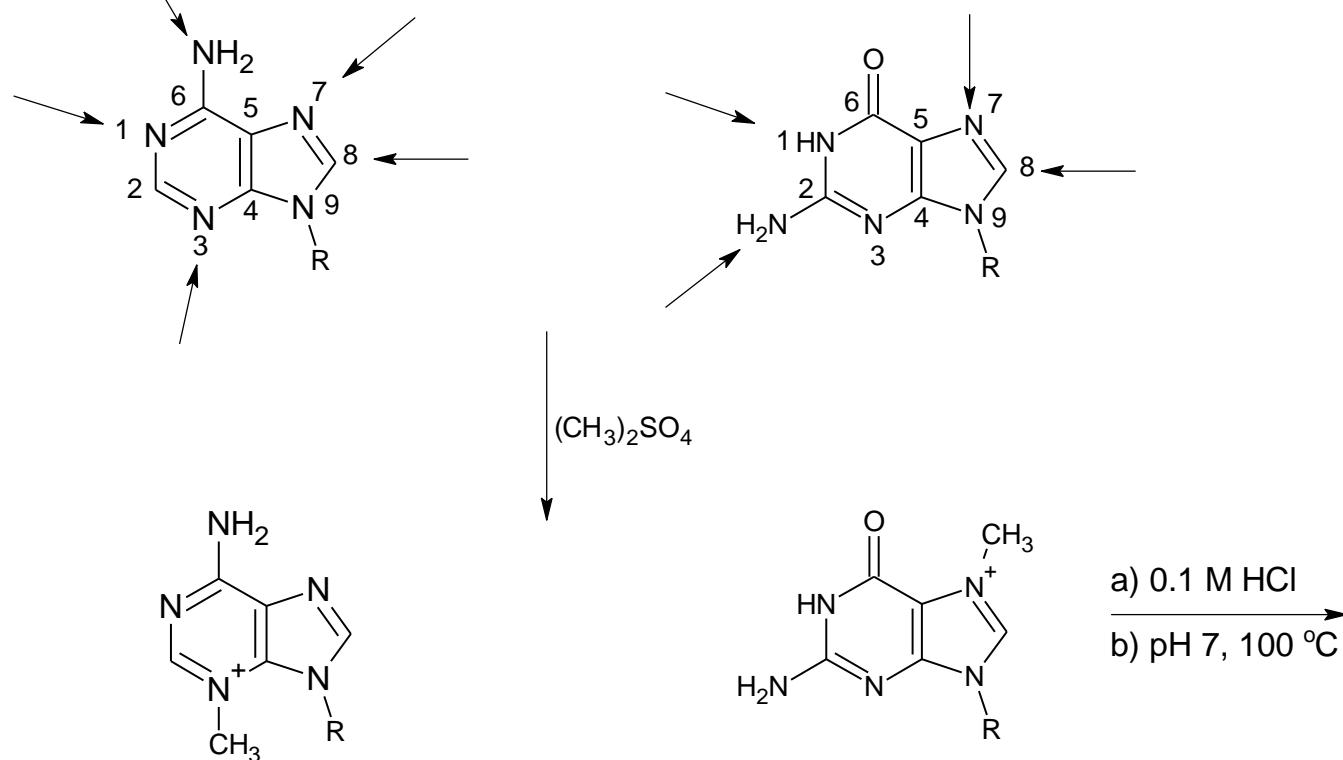
- Periodate oxidation to form oxo-function



oxo-group from (secondary) hydroxyl-function

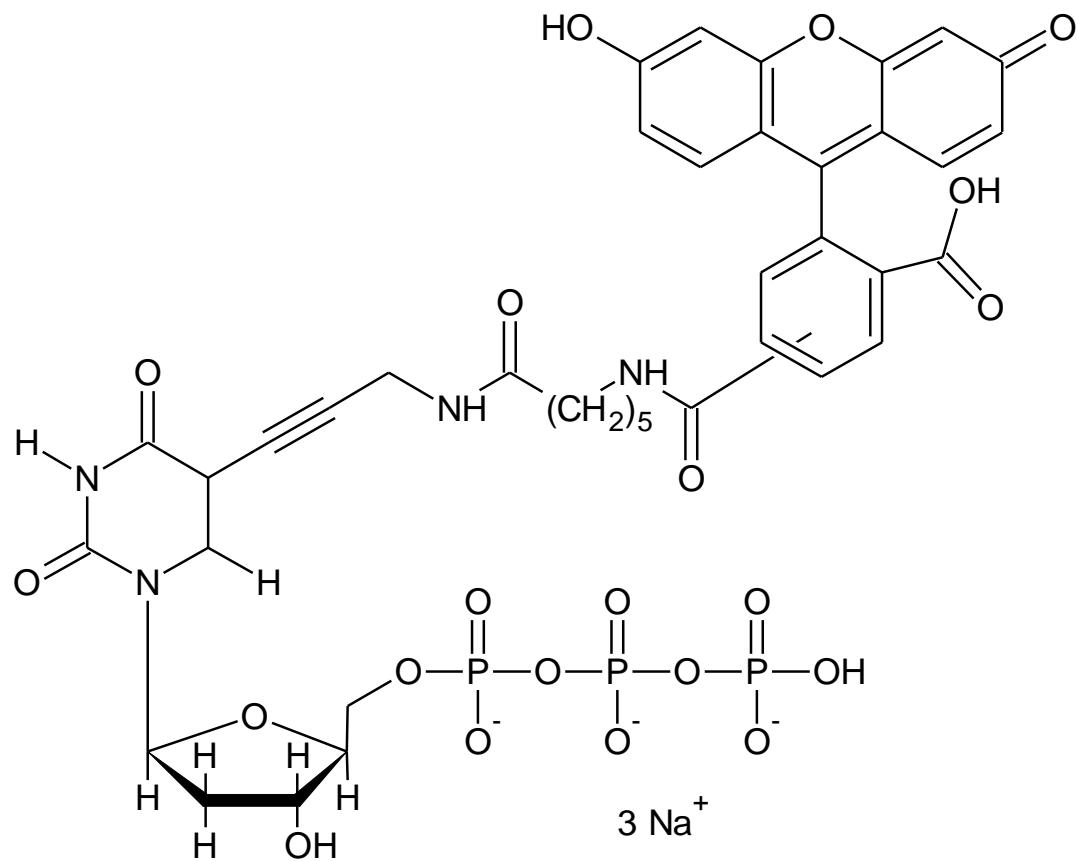
□ Nucleic acids

- Maxam, A.M., Gilbert, W ... et al. A new method for sequencing DNA *PNAS* 74 560-564 (1977)



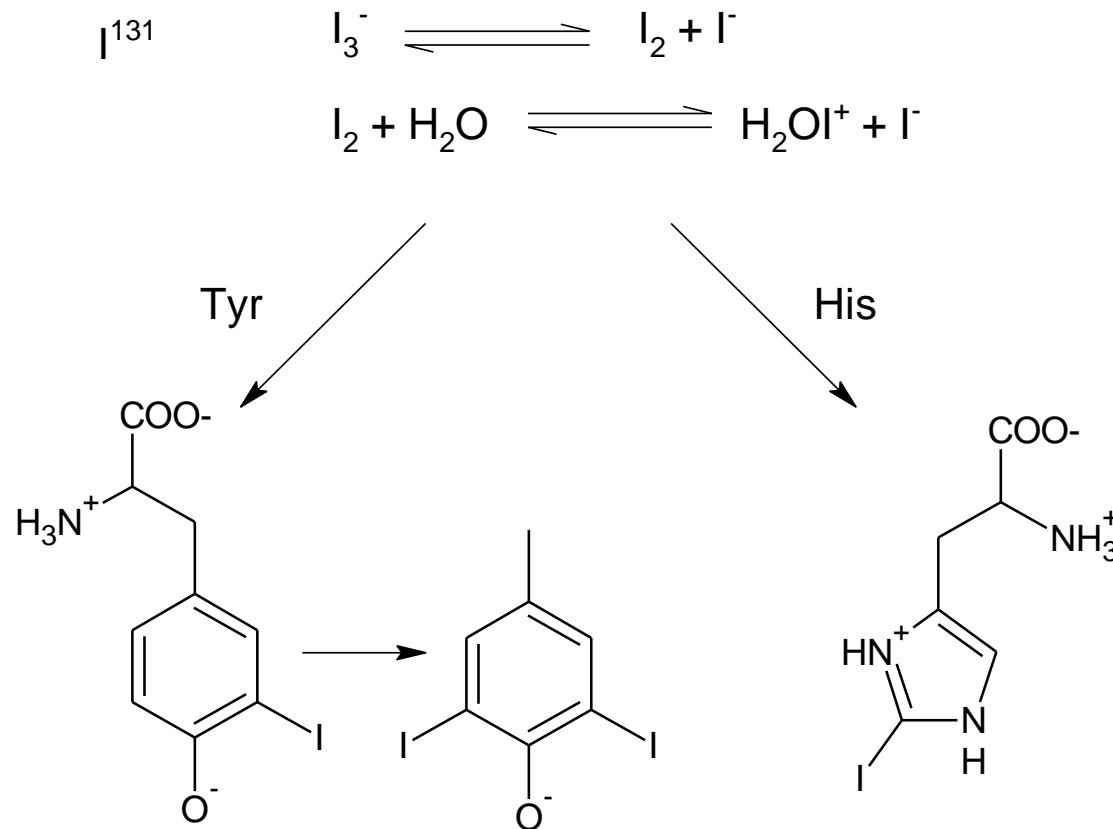
- Sanger, F. et al. *PNAS* 74 5463 (1977)
- Smith, L.M. et al. *Nature* 321 674 (1986)

- Structure of ChromaTide fluorescein-12-dUTP (C-7604)



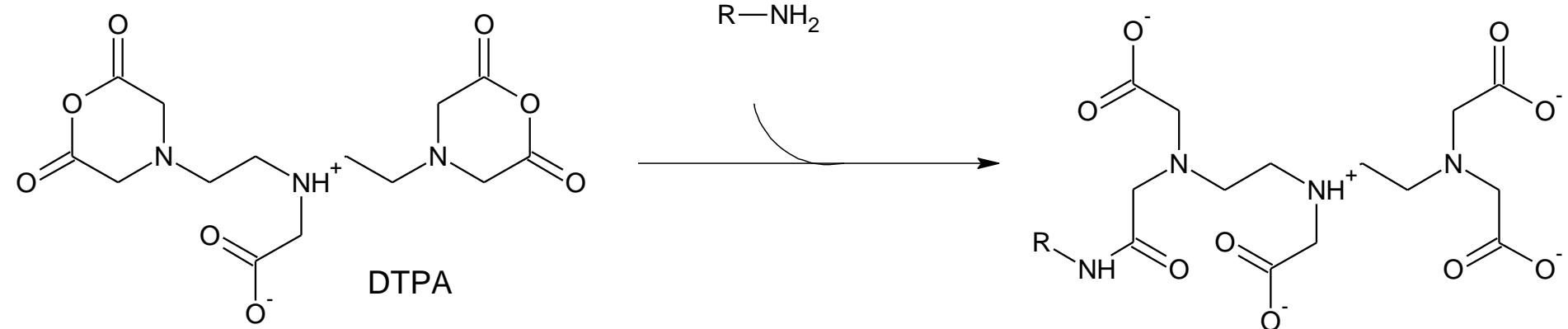
2) Detection of biopolymers in cell (tissue)

- Labelling proteins with radioactive isotope
 - Li, C.H.: Iodination of tyrosine groups in serum albumin and pepsin *JACS* 67 1065 (1945)



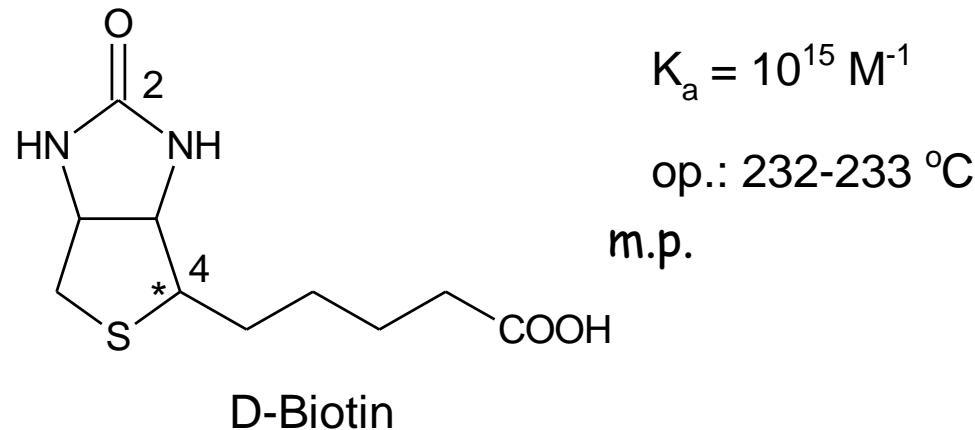
■ Hnatowich, D.J. et al.:

The preparation and labeling of DTPA-coupled albumin *Int J Appl Radiat Isot* 33 327-332 (1982)



□ Labelling of proteins with biotin

- Bayer, E.A., Wilehek, M.: The use of avidin-biotin complex *Methods Biochem Anal* 26 1 (1980)
- Chaiet, L., Wolf, F.J.: The properties of streptavidin, a biotin-binding protein produced by *streptomyces* *Arch Biochem Biophys* 106 1 (1964)
- Green, N.M.: Avidin *Adv Protein Chem* 29 85 (1975)



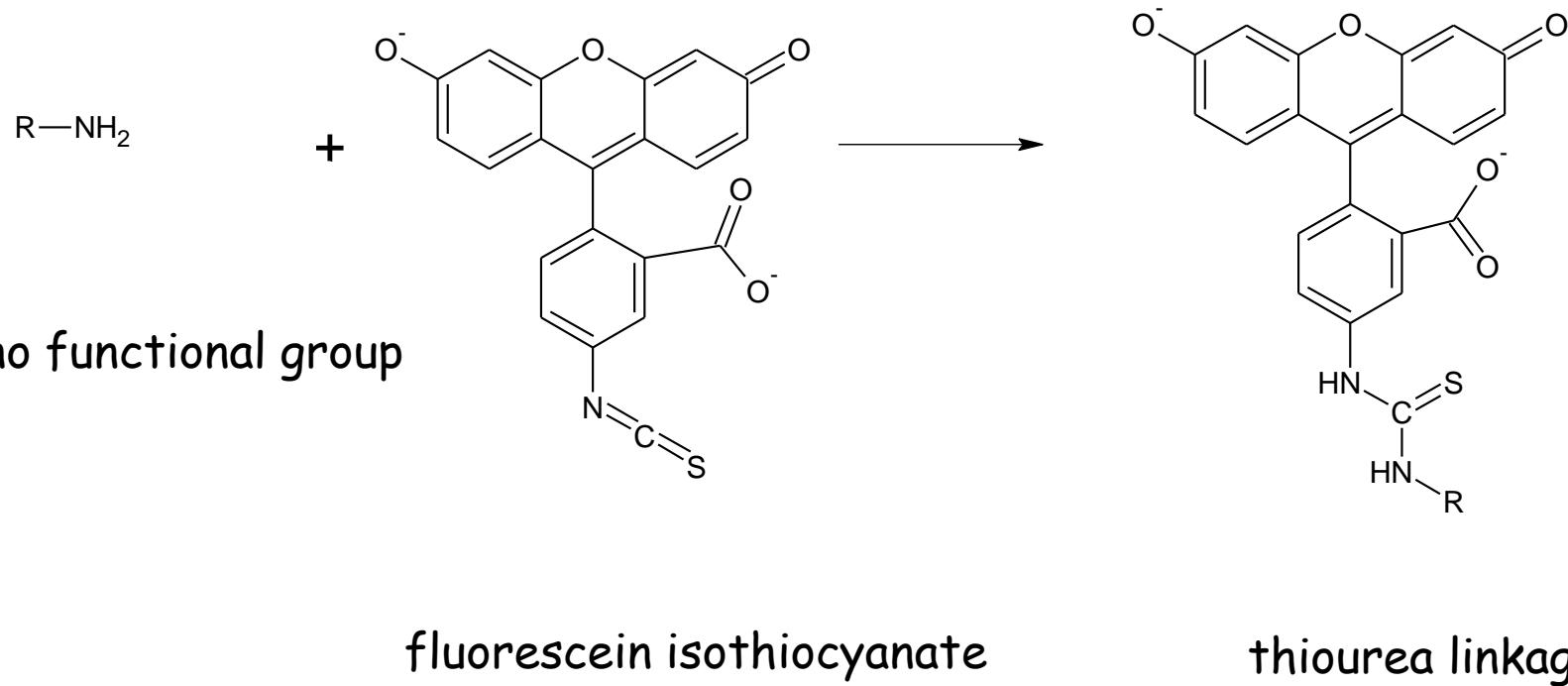
[Hexahidro-2-oxo-1H-tieno[3,4-d]imidazol-4-pentánsav]

Hexahydro-2-oxo-1H-thieno[3,4-d]imidazole-4-pentanoic acid; (+)-Biotin

□ Labelling of biopolymers with fluorophore

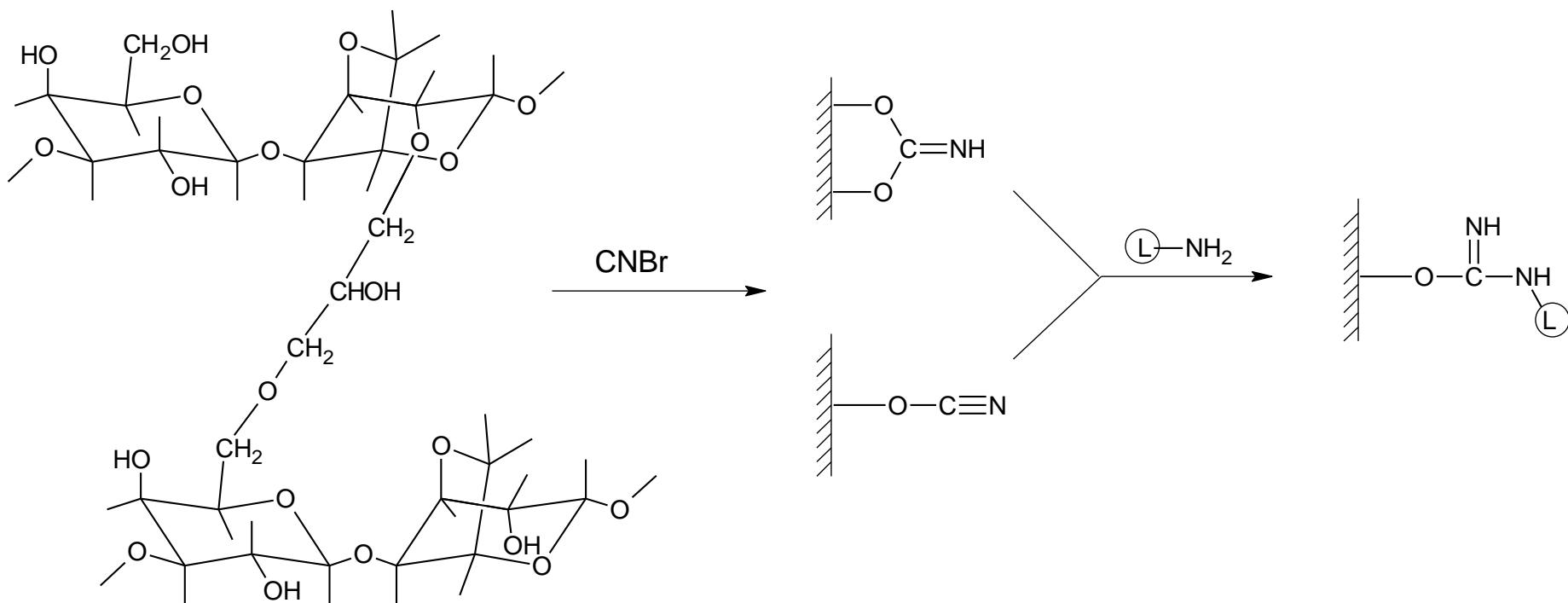
McKinney, R. et al.

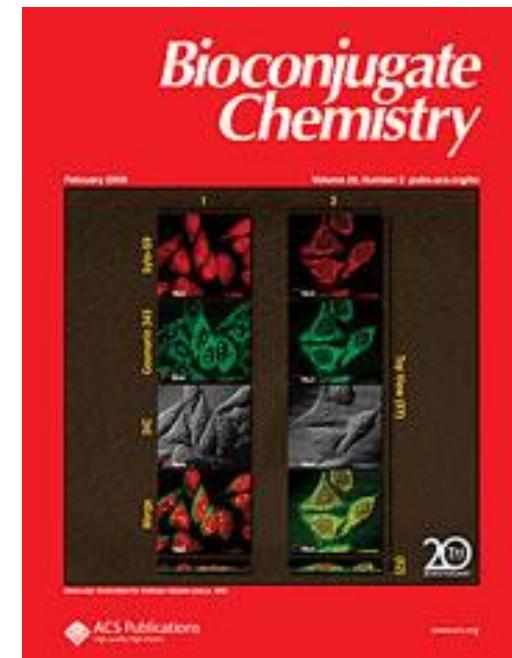
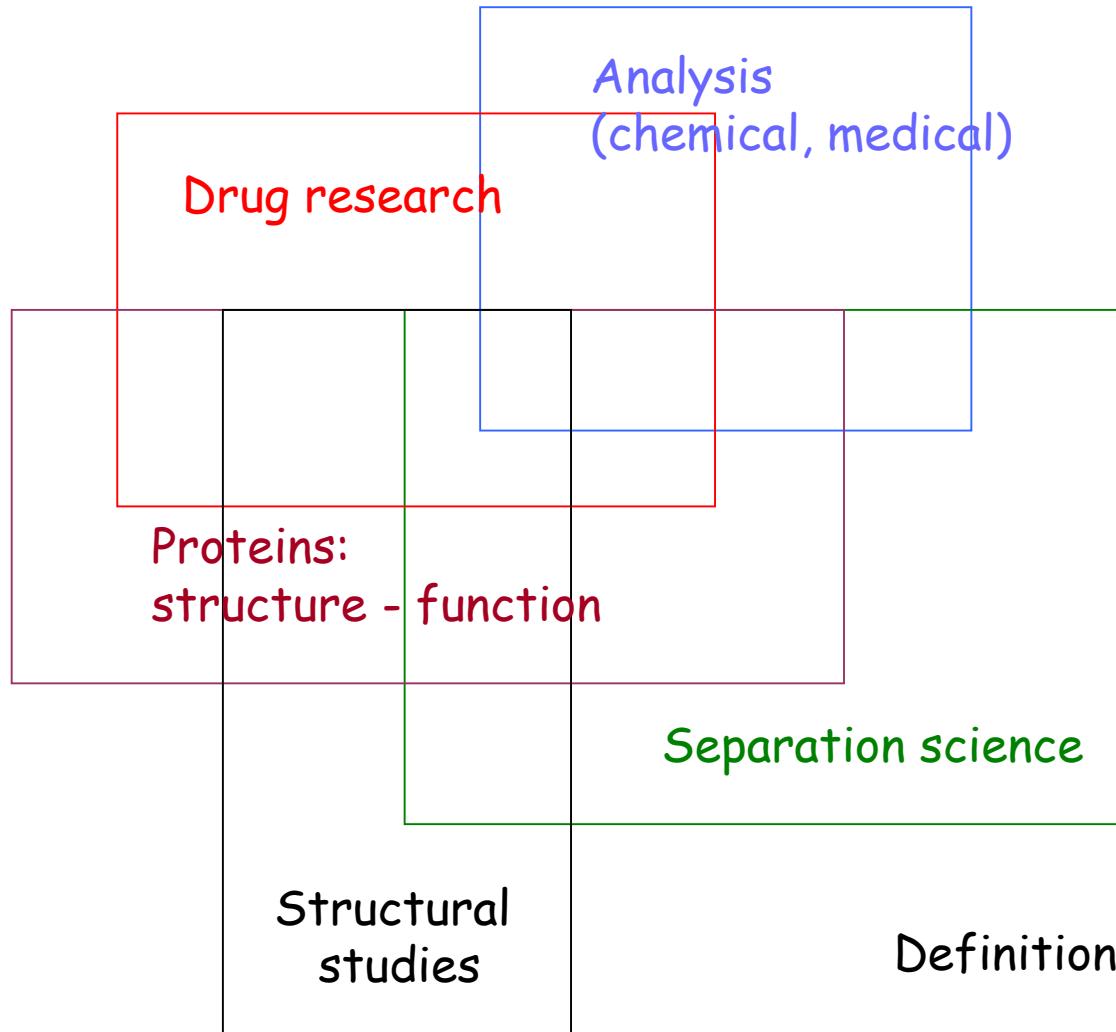
Factors affecting the rate of reaction of fluorescein isothiocyanate with serum proteins *J Immunol* 93 232 (1964)



2) Affinity chromatography

- Bethell, G.S. et al. A novel method of activation of cross-linked agarose with 1,1'-carbonyldiimidazol which gives a matrix for affinity chromatography
J Biol Chem 254 2572 (1979)





Definitions

1. Two or more active components
2. Covalent linkage

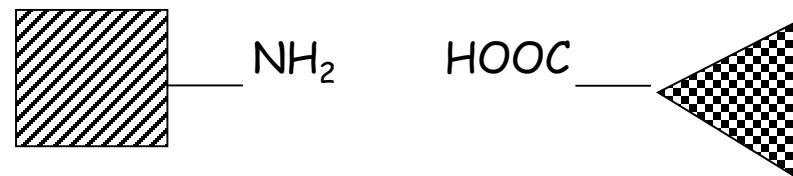
■ Approaches

1. Size of the partners

- Small - small
- Small - big
- Big - big

2. Type of linkage

- Direct

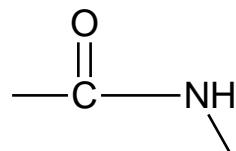


- Indirect

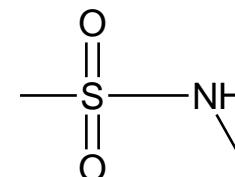


3. Type of linkage

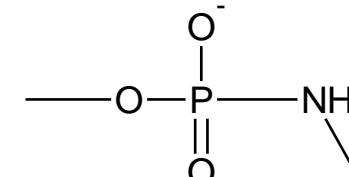
- Acid amide



carboxylic acid

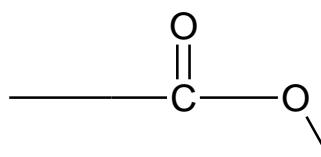


sulphonic acid

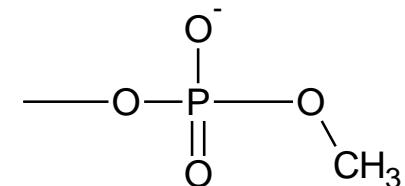


phosphoric acid amid

- Acid ester

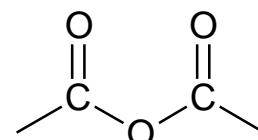


carboxylic acid

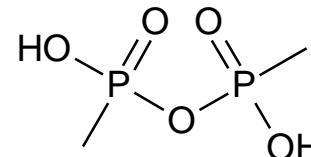


phosphoric acid

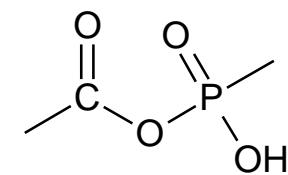
- Acid anhydride



carboxylic acid

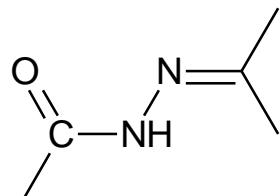


phosphoric acid



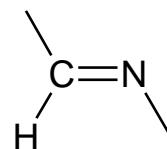
mixed

- Hydrazone



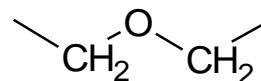
Aldehyde + hydrazine

- Schiff base



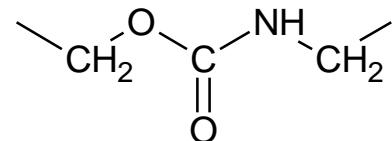
Aldehyde + amine

- Ether



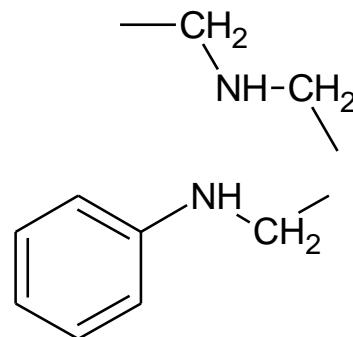
Alcohol

- Carbamate



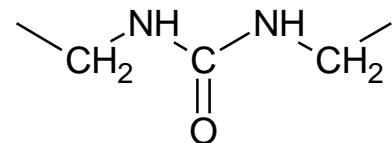
Alcohol

- Secunder amine
(N-glycoside)



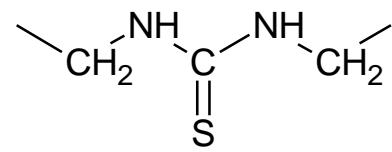
Amine
+ aldehyde
+ aryl-halogenide

- Isourea



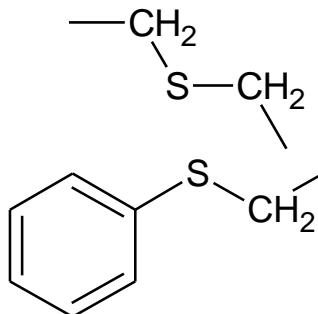
Amine

- Isothiourea



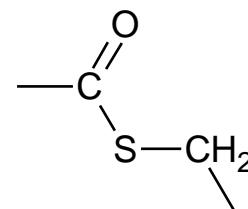
Amine

- Thioether



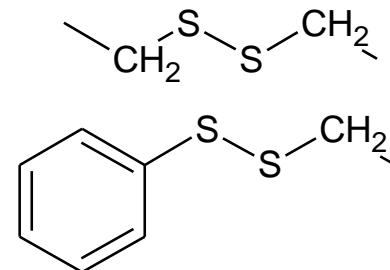
Thiol

- Thioester



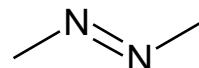
Thiol

- Disulphide



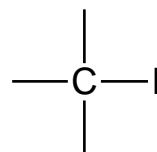
Thiol

- Diazo



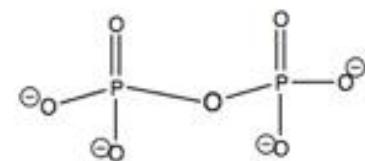
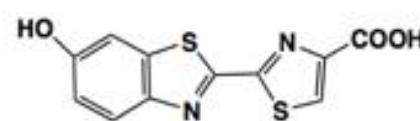
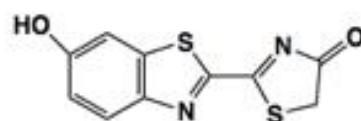
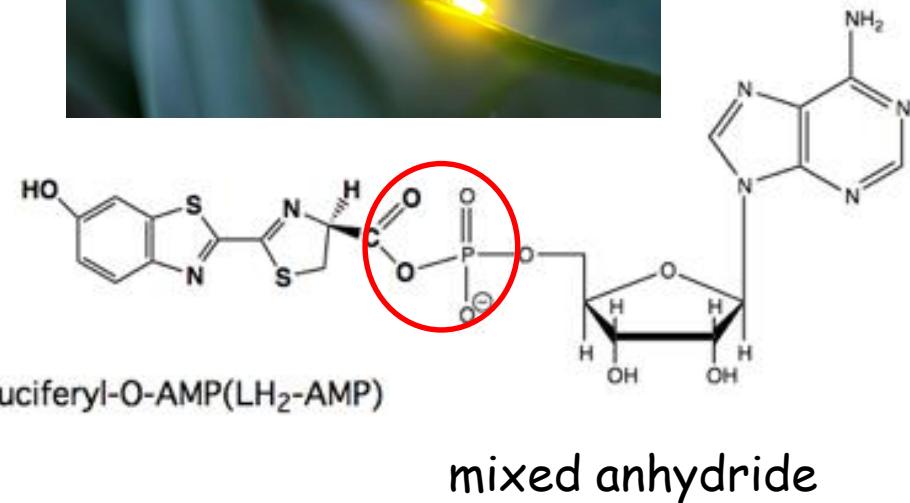
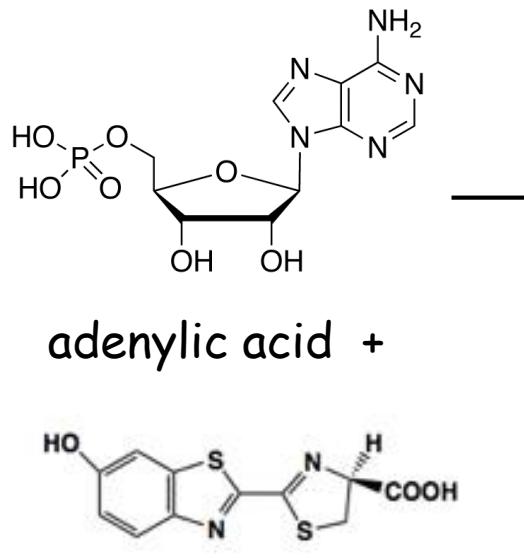
Azide

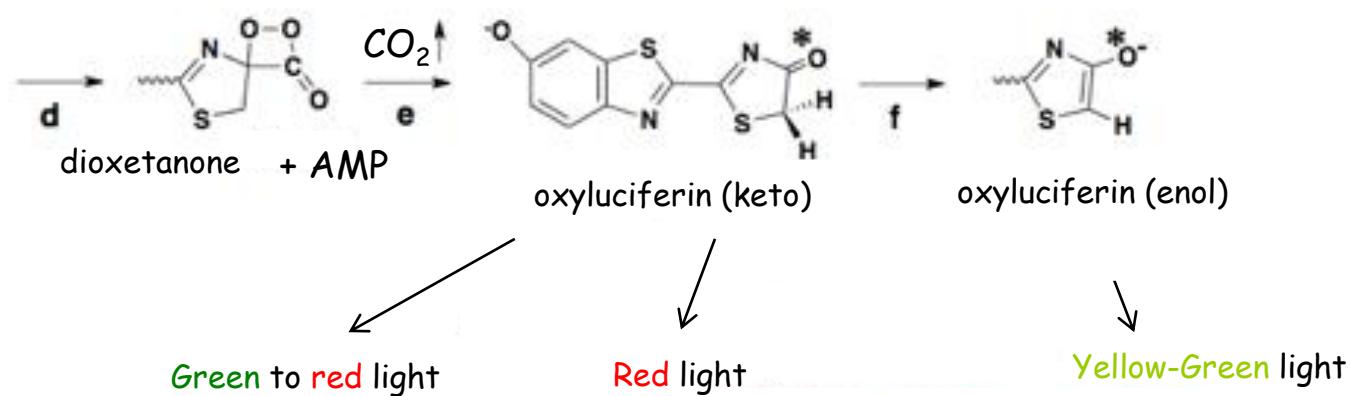
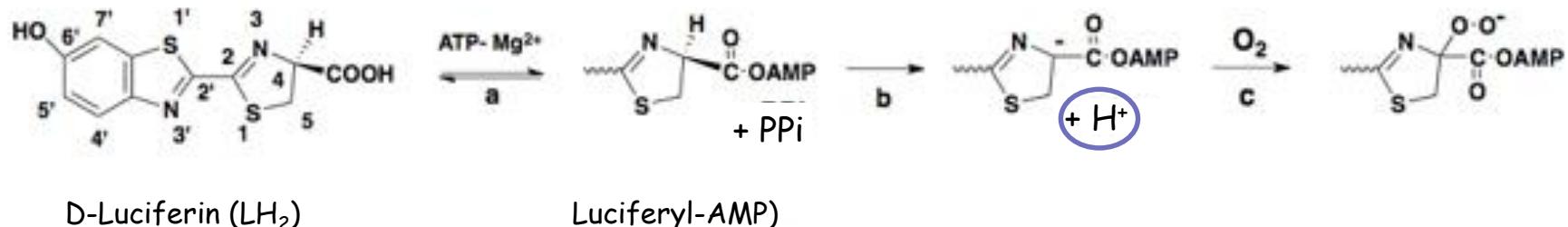
- $C - X$



1. Example: small-small, direct, anhydride bond

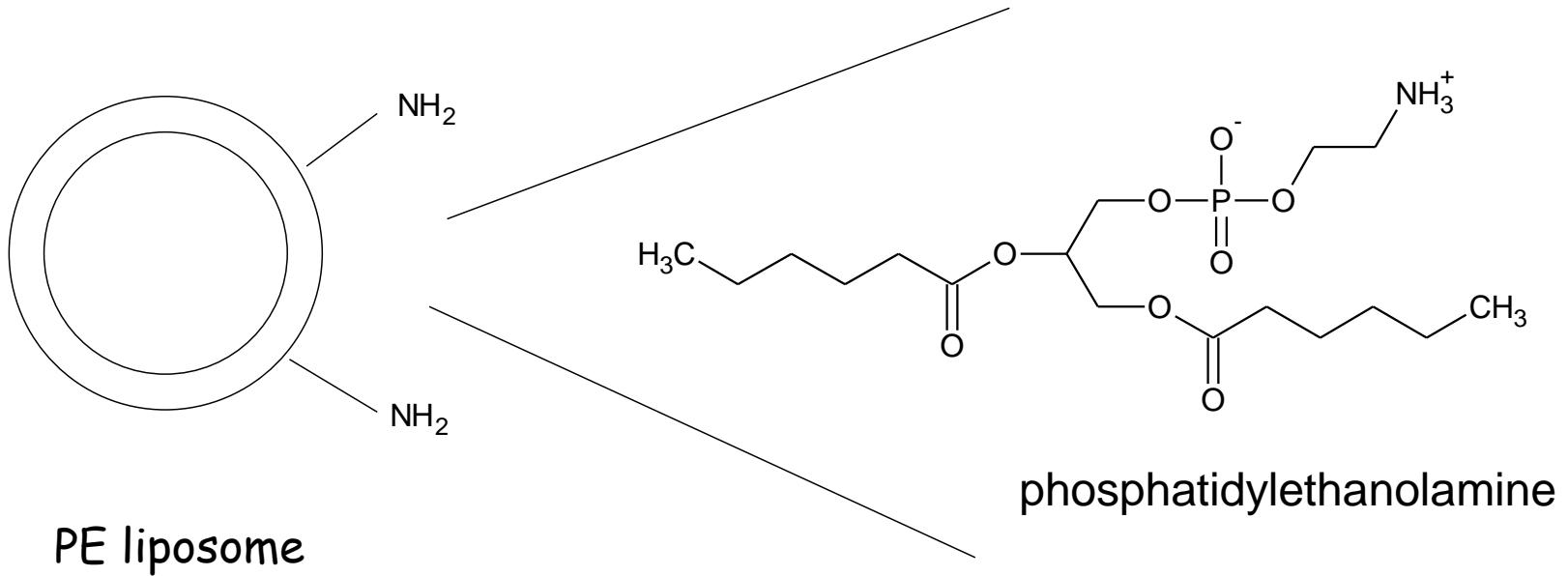
„Firefly“ luminescence



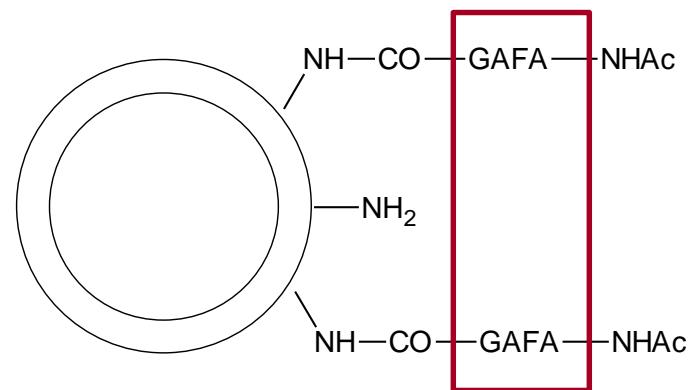
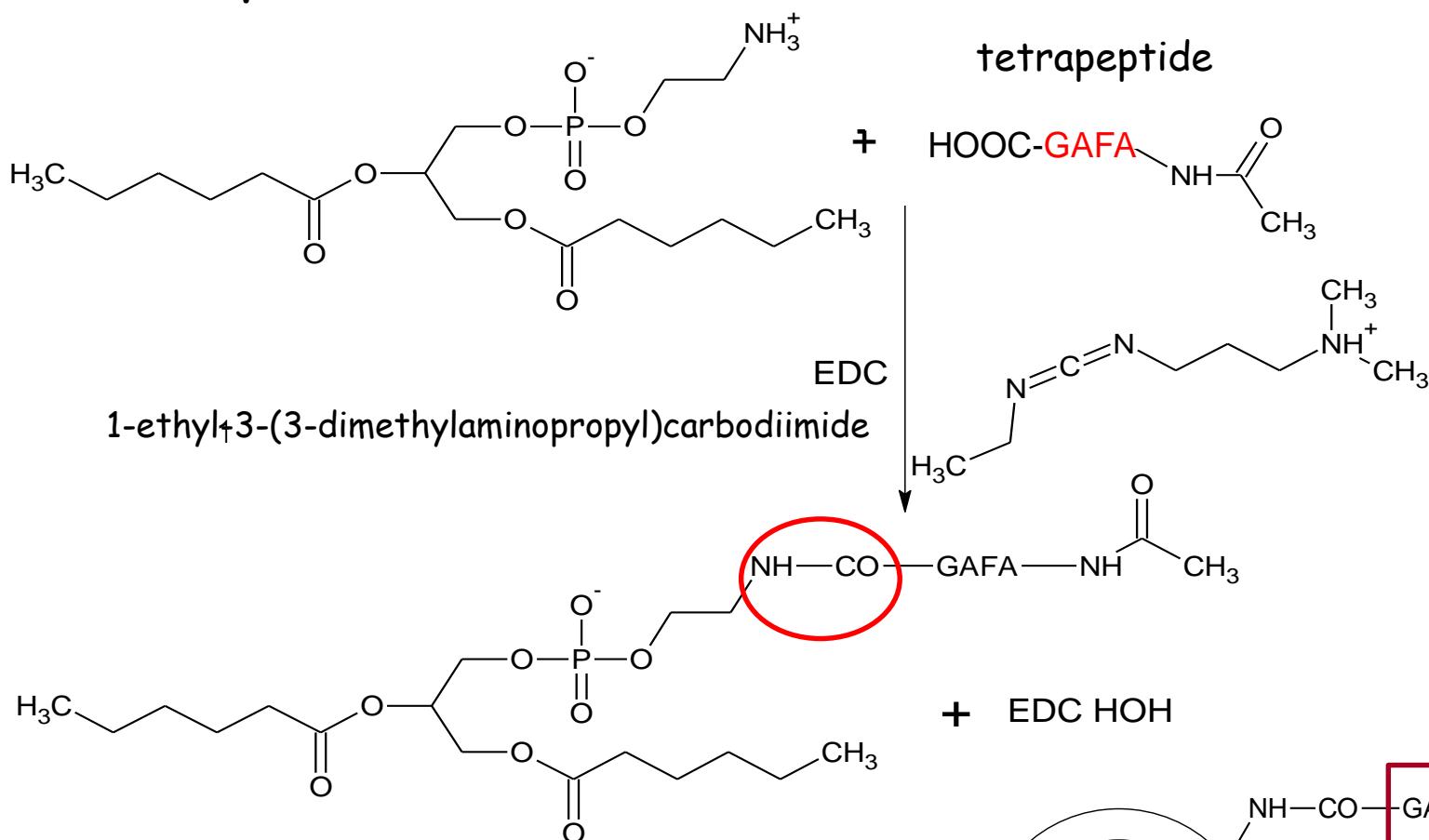


- Step a: formation of the enzyme-bound luciferyl adenylate;
- Step b: a proton is abstracted from the C-4 carbon of the adenylate by a basic side chain amino acid residue of **luciferase**;
- Step c: molecular oxygen adds to the newly formed anion;
- Step d: a highly reactive dioxetanone intermediate is formed;
- Step e: an electronically excited state oxyluciferin molecule, CO_2 and red light emission ($\lambda_{max} = 615\text{ nm}$) are produced (at pH 6);
- Step f: the enolate dianion formed by **tautomerization** from the exited form of oxyluciferin and yellow-green light emission ($\lambda_{max} = 560\text{ nm}$) occurs at pH 8.

2. Example: small - big, indirect, amide bond
Liposome - hapten conjugate



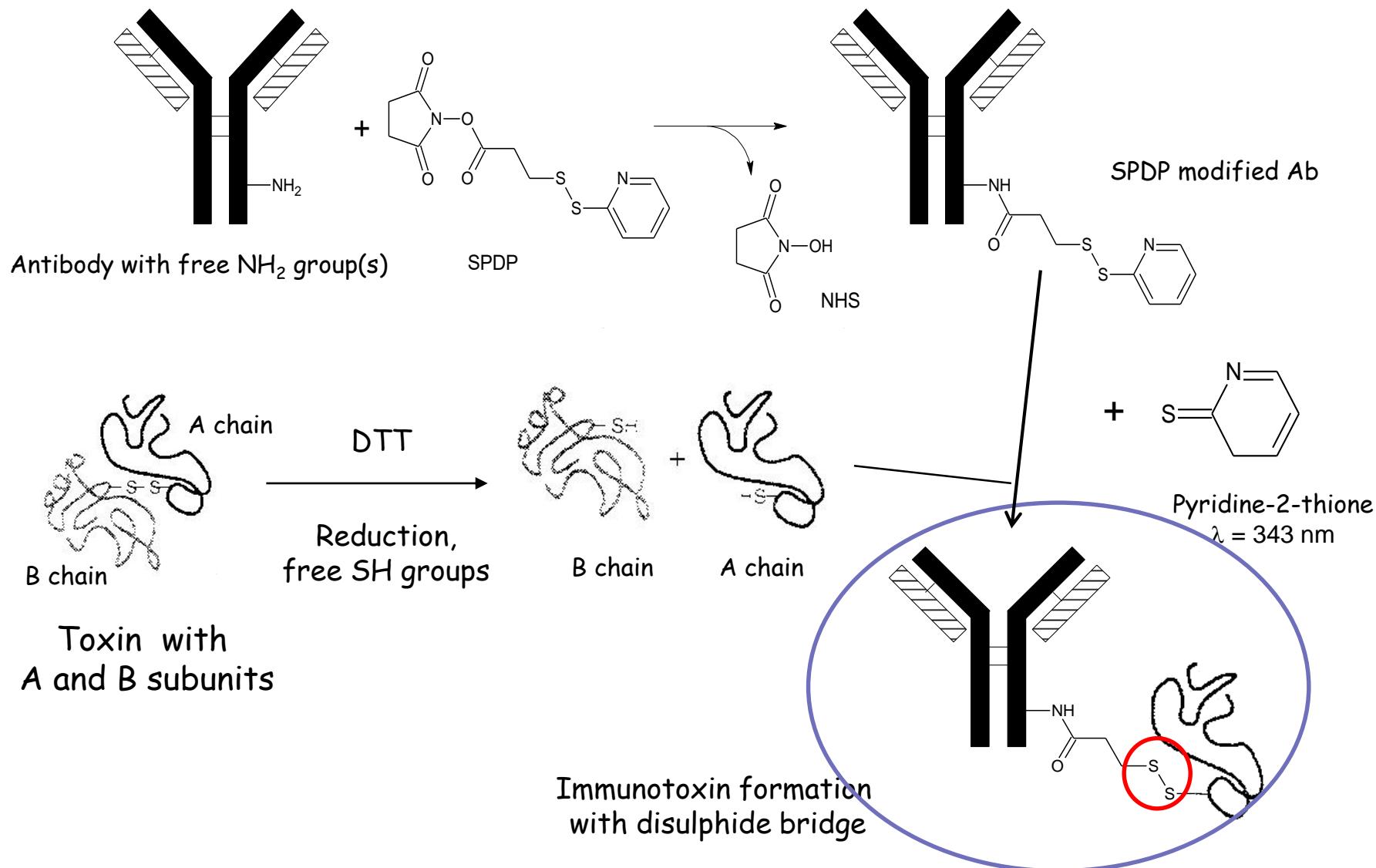
2. Example



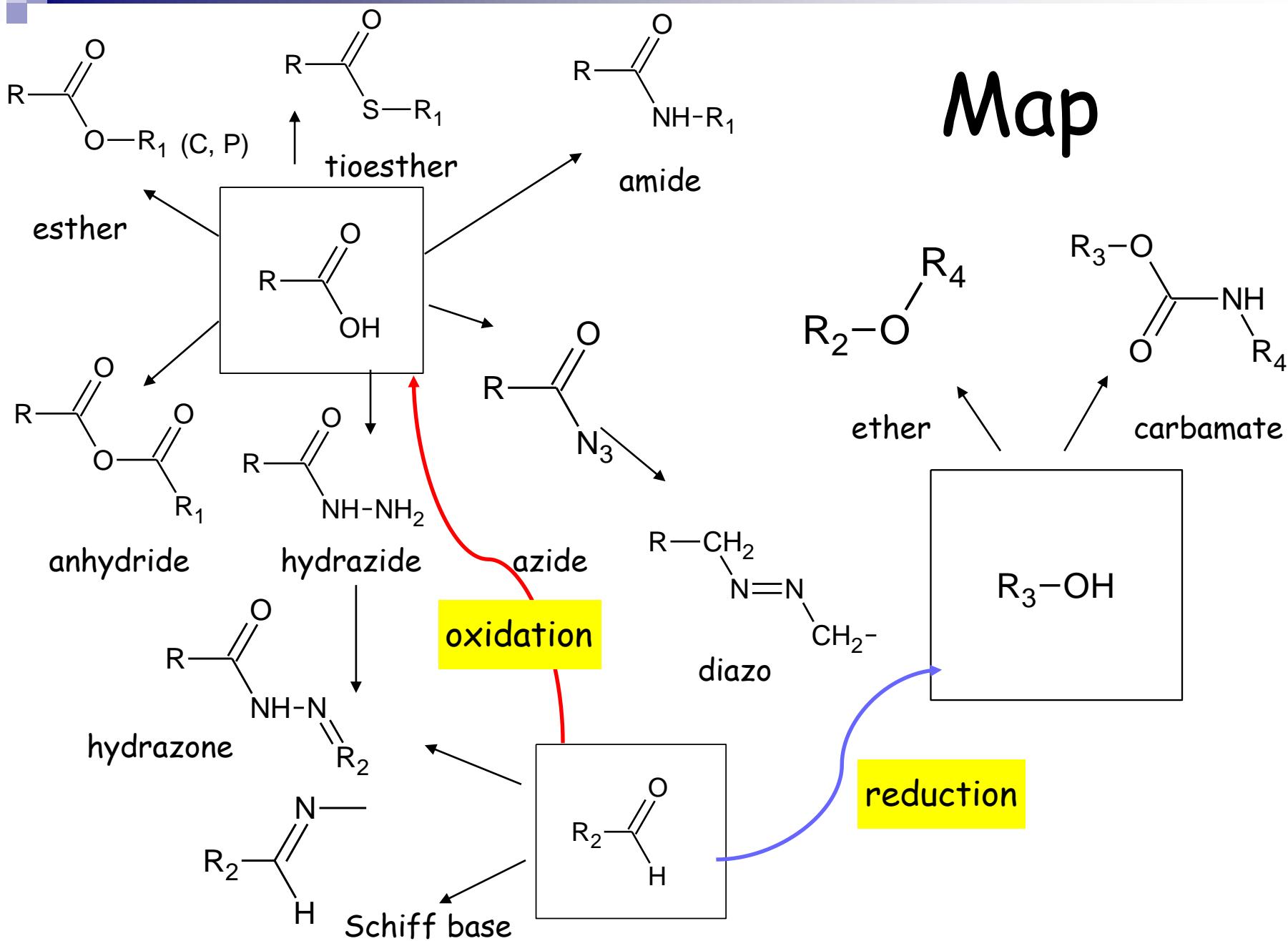
3. Example: big-big, indirect, disulphide bond
Immunotoxin conjugate
1. step: introduction of protected -SH group(s) of antibody ($\text{Ab} + \text{SPDP} \rightarrow$ amide bond)
 2. step: creation of free -SH groups of the toxin subunits (cleavage of the -S-S- bond, DTT)
 3. step: free -SH partner reacts with the component having „protected” SH function ($\text{AB-SSP} + \text{Toxin-SH} \rightarrow$ disulphid linkage)

3. Example

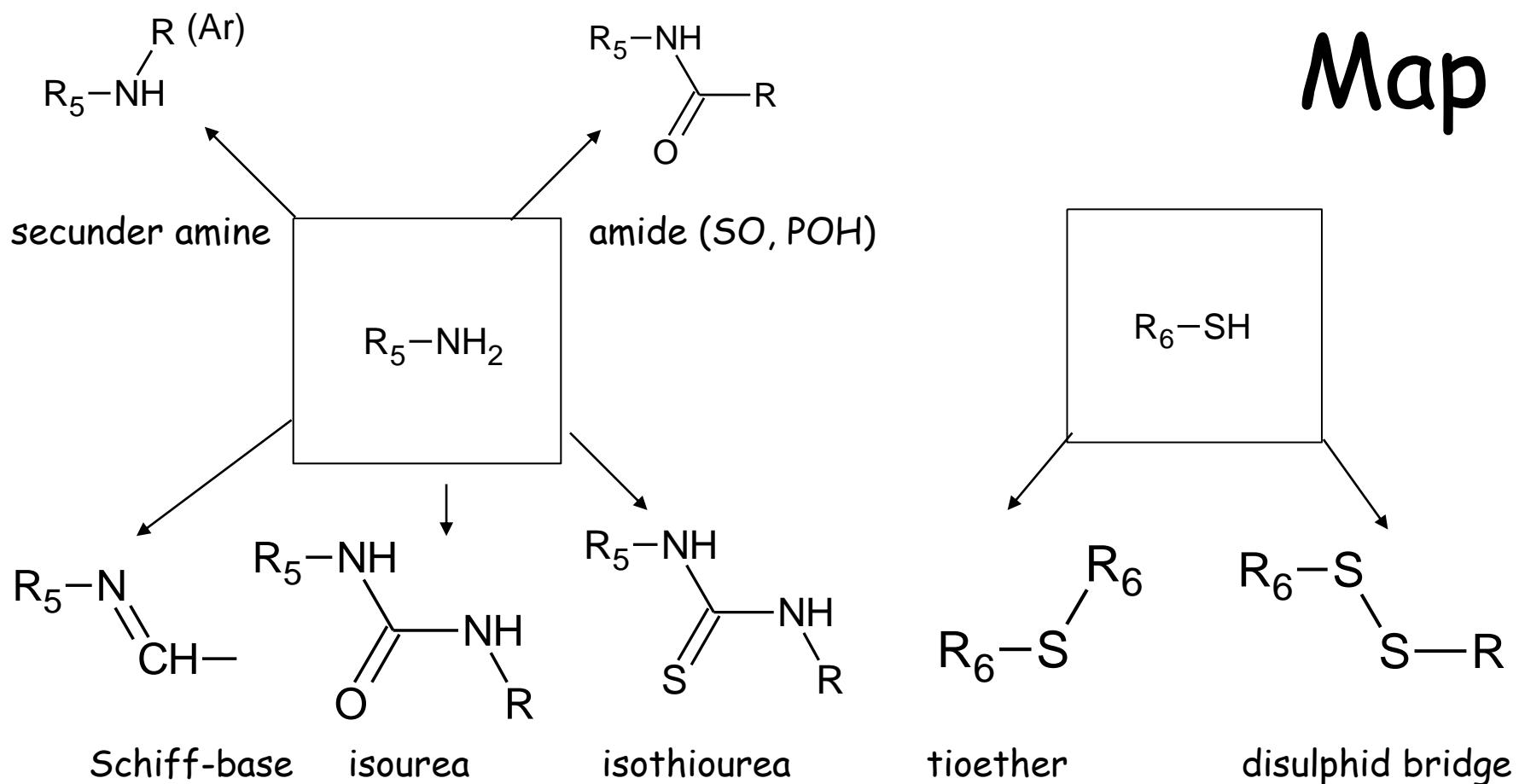
□ Carlsson, J. et al. Biochem J 173 723 (1978)



Map



Map



- 1) Introduction of functional groups? (-COOH, -CHO, -OH, -SH, -NH₂)
- 2) Elimination of functional groups?